

# Extraction of Secondary Metabolites from *Mimosa pudica* L. and Study of its Antimicrobial Activity

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Abstract— Now-a- days medicinal plants have high medicinal values and economic importance. The secondary metabolites are mainly involved in the protection but these are not necessary for a cell to live. During the adverse condition, the secondary metabolites are secreting from the plants to protect them from the infection of bacteria and insects. The present investigation emphasized on the extraction and antimicrobial potential of Mimosa pudica (M. pudica) plants. These plants are highly riched with alkaloids, flavonoids, terpene etc. They have great economic medicinal value due to its use in the treatment diabetes, bleeding piles, hepatitis, cancer etc. Here we collected the plants from GIET University campus and extracted the secondary metabolites from its leaf and investigated the antimicrobial potential against E. coli with different concentration of aqueous extract. We found the significant antimicrobial activity against E. coli. The diameter of inhibition zone was found 15mm, 13 mm, 12mm and 11 mm with concentration of 25µl, 50µl, 75µl and 100µl of extract respectively. This result confirms that M. pudicais possessing the tremendous antimicrobial activity against the E. coli bacteria.

*Keywords*— Seondary metabolites, Mimosa pudica, Antimicrobial activity, Zone of inhibition, Aqueous extract

## I. INTRODUCTION

From ancient times, plants contributed valuable sources of natural products. In recent times plants are having different biological activities like antimicrobial, antiviral, antivenom, antioxidant used widely in pharmaceutical and food industries [1]. According to the WHO, traditional medicine was used by around 80% population of world. As per several research studies, it is seen that medicine derived from plants has low side effect on human health than synthetic substances [2]. The antimicrobial properties of compound extracted from medicinal plant have showing to be extremely important in medical field applications. These compounds such as flavonoids, terpenes, alkaloids, tannins, and essential oils are synthesized by the plants secondary metabolism. Alkaloids are natural, organic substance found mostly in plants that have at least one nitrogen atom in their chemical structure. Alkaloids act

as a defence against animal or insect or bacteria [3]. Alkaloids are found a variety of plants including M. Pudica L. M. pudica L. is a perennial herb, compound leaves. It shows thigmonastic movement [4]. In some plant (Mimosa species) herbaceous to woody plant. M. Pudica diffuse under shrubs, the height of plant is 50-90cm. Mimosa pudica is a sensitive plant generally called as touch me not plant. It is closing its leaves when it will be touched. In behaving this way M. pudica demonstrate thigmonasty [4]. The turgor pressure of the cells causes the bending of Mimosa pudica leaves. Except seisomonastic movements, this plant also closes its leaves under higher temperature. The plant bends inward due to a series of bioelectrical and biochemical changes occur under external stimuli. The movement of leaves and stems starts at pulvinus which is a swelling pad like petioles. Pulvins contain two types of cells called as extensor cells and flexor cells which are present oppositely. Under physical touch, stretch occurs in flexor cells while the extension occurs in extensor cells. The turbor pressure of extensor cells cause the bending [4, 5]. For healthcare purpose many developing and developed countries are using plants and natural products. M. pudica leaves have antimicrobial, anticancer, anti diuretic actions. It helps the gut by absorbing toxin, heavy metals, parasites, bacteria. It facilities the excretions of the body remove the toxin material from the body. It's keep the digesting process less pressure and maintaining clean and health gut. The principle of brain and gut relationship really want to know natural weight maintain the mental status [6, 7, 8]. 90 percent of serotine made in gut. It is the happy hormone that is responsible for maintaining mood. Bacteria in the gut has the capability to help the body produces serotine. Lack of neurotransmitters is associate certain health problems such as anxiety, depression [9]. M. pudica has the caper to maintain gut health by removing body waste and maintain good bacteria growth. Plants are the major source of phytomedicine for human health care and it is prudent to analyse the various phytoconstituents present in them. Phytochemical analysis of Mimosa pudica L. showed presence of alkaloids, flavonoids, phenols [4, 10]. Hence, the current work emphasized on the extraction of secondary metabolites from Mimosa pudica and investigated its antimicrobial activity.

MATERIALS AND METHODOLOGY PLANT MATERIAL COLLECTION

The plant materials were collected from GIET University campus and transported to Plant Science laboratory and properly washed with tap water. After the washing the plant materials were dried under normal temperature for two days and again expose to oven drying at 70°C for 3 hours. The dried leaves were grinded into powder form and subjected to

aqueous extraction.

## AQUEOUS EXTRACTION

Two grams of leaf powder was taken and dissolved in 100ml of distilled water and incubated for 48 hours. Then the sample was further filtered by Whatman no.1 filter paper to discard the cell debris. The filtered supernatant was collected in a test tube [11].

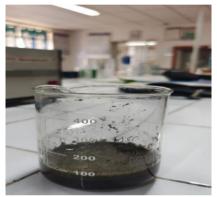


Fig:1 Aqueous extract from Mimosa pudica L

## SOLVENT EXTRACTION

Two grams of powdered leaves were taken for solvent extractions using Soxhlet apparatus. Here ethanol used as solvent. 150ml of solvent was prepared by adding 75% ethanol with 25% distilled water (112.5 ethanol and 37.5 ml distilled water) [12, 13].



Fig:2 Solvent extract from Mimosa pudica L

### PROCESS OF SOXHELT EXTRACTION

The solvent taken in Soxhelt apparatus for assembling and solvent is heated under reflex. Solutes are taken and transferred from the extraction chamber into the reservoir. Reflex it for 5 cycles. After completion of each cycle the fresh solvent is extracted and collected in the Soxhelt apparatus. At last the extracted sample is collected in a centrifuge tube [12].



Fig:3 Soxhelt extraction from Mimosa pudica L

#### ANTIMICROBIAL ACTIVITY

The antimicrobial activity of *M pudica* plant's extract against *E. coli* is determined by agar well diffusion method. The aqueous extract taken to study the antimicrobial properties of *M. pudica*. In the very first step, we have taken 1.4 gram of nutrient agar and added 50ml of distilled water inside a beaker. Then petridish, conical flask with distilled water and agar is taken to autoclave for sterilization. After this process, all the sterilized materials were transferred into the laminar air flow to prevent from getting contaminated [14].

Then nutrient agar is taken in a petridish and it allowed to drt for 15 minutes. Then a sterilized cork borer was used to make well. The *E. coli* bacteria collected from MTCC were spread on solid agar plate using a spreader. Variable concentrations (25, 50, 75 and 100µl) of plant extract test sample were added into the every well. The plates were labelled and kept for incubation at 37°C overnight. Then zone of inhibition zone was calculated and diameters of zones were noted [15].

## **RESULTS AND DISCUSSION**

The antimicrobial activity of *M. Pudica* tested against gram negative bacteria *E. coli*. It was found that aqueous extract of *M. pudica* confirmed antimicrobial potential and shown the most effective inhibition zone against *E. coli*.

We found that the inhibition zone was increased when the concentration of aqueous extract is decreased. The difference in the observe zone of inhibition activity is due to the varying degree of solubility of the active constituent as in extract.



Fig:4

Treatment

of

aqueous from *Mimosa pudica* L against *E. Coli* by agar diffusion method

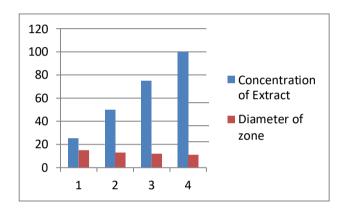


Fig: 5 Comparative analysis of zone of inhibition in different concentration

Sl no.	Concentration of Extract	Diameter of zone	Results
1	25	15mm	+
2	50	13mm	+
3	75	12mm	+
4	100	11mm	+

Table:1 Zone of Inhibition of Mimosa pudica L against E. coli

# CONCLUSION

The current research work concluded that the antimicrobial activity of the *M. pudica* plant extract would be more effective against *E. coli* bacteria and the disease associated by *E. coli*. Also it is found that the aqueous extract have antagonistic effects on broad spectrum gram -ve bacteria. Further research work can be conducted on broad range of bacteria to see more antagonistic effects. Also investigation on identification and purification of bioactive compounds associated with *M. pudica* and their toxicological effects can be investigated for the development of future potential drugs for human welfare.

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## **REFERENCES:**

- Alhazmi HA, Najmi A, Javed SA, Sultana S, Al Bratty M, Makeen HA, Meraya AM, Ahsan W, Mohan S, Taha MME, Khalid A. Medicinal Plants and Isolated Molecules Demonstrating Immunomodulation Activity as Potential Alternative Therapies for Viral Diseases Including COVID-19. Front Immunol. 2021 May 13;12:637553. doi: 10.3389/fimmu.2021.637553
- Ekor M. The growing use of herbal medicines: issues relating to adverse reactions and challenges in monitoring safety. Front Pharmacol. 2014 Jan 10;4:177. doi: 10.3389/fphar.2013.00177.
- Othman L, Sleiman A, Abdel-Massih RM. Antimicrobial Activity of Polyphenols and Alkaloids in Middle Eastern Plants. Front Microbiol. 2019 May 15;10:911. doi: 10.3389/fmicb.2019.00911.
- Ahmad H, Sehgal S, Mishra A, Gupta R. Mimosa pudica L. (Laajvanti): An overview. Pharmacogn Rev. 2012 Jul;6(12):115-24. doi: 10.4103/0973-7847.99945.
- N. R. Deepak and S. Balaji, "Performance analysis of MIMO-based transmission techniques for image quality in 4G wireless network," 2015 IEEE

International Conference on Computational Intelligence and Computing Research (ICCIC), 2015, pp. 1-5, doi: 10.1109/ICCIC.2015.7435774.

 De Luccia TP, Friedman P. Boolean function applied to Mimosa pudica movements. Plant Signal Behav. 2011 Sep;6(9):1361-4. doi: 10.4161/psb.6.9.16445.

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- T. N and D. N R, "A Convenient Machine Learning Model for Cyber Security," 2021 5th International Conference on Computing Methodologies and Communication (ICCMC), 2021, pp. 284-290, doi: 10.1109/ICCMC51019.2021.9418051.
- Mahomoodally MF. Traditional medicines in Africa: an appraisal of ten potent african medicinal plants. Evid Based Complement Alternat Med. 2013;2013:617459. doi: 10.1155/2013/617459. Epub 2013 Dec 3.
- Alamgir ANM. Biotechnology, In Vitro Production of Natural Bioactive Compounds, Herbal Preparation, and Disease Management (Treatment and Prevention). Therapeutic Use of Medicinal Plants and their Extracts: Volume 2. 2018 Jun 24;74:585–664. doi: 10.1007/978-3-319-92387-1\_7.
- N R, Deepak and G K, Dr. Suhas and B, Bhagappa and Kumar Pareek, Piyush, A Framework for Food recognition and predicting its Nutritional value through Convolution neural network (February 22, 2022). Available at SSRN: <u>https://ssrn.com/abstract=4040968</u> or <u>http://dx</u>. <u>doi.org/10.2139/ssrn.4040968</u>.
- Prasathkumar, M., Anisha, S., Dhrisya, C., Becky, R., & Sadhasivam, S. (2021). Therapeutic and pharmacological efficacy of selective Indian medicinal plants – A review. Phytomedicine Plus, 1(2), 100029. doi:10.1016/j.phyplu.2021.100029
- K. M P and D. N R, "Crop Prediction Based on Influencing Parameters for Different States in India-The Data Mining Approach," 2021 5th International Conference on Intelligent Computing and Control Systems (ICICCS), 2021, pp. 1785-1791, doi: 10.1109/ICICCS51141.2021.9432247.
- Clapp M, Aurora N, Herrera L, Bhatia M, Wilen E, Wakefield S. Gut microbiota's effect on mental health: The gut-brain axis. Clin Pract. 2017 Sep 15;7(4):987. doi: 10.4081/cp.2017.987.
- Oikeh EI, Omoregie ES, Oviasogie FE, Oriakhi K. Phytochemical, antimicrobial, and antioxidant activities of different citrus juice concentrates. Food Sci Nutr. 2015 Jul 30;4(1):103-9. doi: 10.1002/fsn3.268.
- 15. Umamaheswari A, Prabu SL, John SA, Puratchikody A. Green synthesis of zinc oxide nanoparticles using leaf extracts of *Raphanus sativus var*. *Longipinnatus* and evaluation of their anticancer

property in A549 cell lines. Biotechnol Rep (Amst). 2021 Feb 5;29:e00595. doi: 10.1016/j.btre.2021.e00595.

- Redfern J, Kinninmonth M, Burdass D, Verran J. Using soxhlet ethanol extraction to produce and test plant material (essential oils) for their antimicrobial properties. J Microbiol Biol Educ. 2014 May 1;15(1):45-6. doi: 10.1128/jmbe.v15i1.656.
- Zhang QW, Lin LG, Ye WC. Techniques for extraction and isolation of natural products: a comprehensive review. Chin Med. 2018 Apr 17;13:20. doi: 10.1186/s13020-018-0177-x.
- Deepak, N.R., Balaji, S. (2016). Uplink Channel Performance and Implementation of Software for Image Communication in 4G Network. In: Silhavy, R., Senkerik, R., Oplatkova, Z., Silhavy, P., Prokopova, Z. (eds) Software Engineering Perspectives and Application in Intelligent Systems. CSOC 2016. Advances in Intelligent Systems and Computing, vol 465. Springer, Cham. <u>https://doi.org/10.1007/978-3-319-33622-0\_10</u>
- Gonelimali FD, Lin J, Miao W, Xuan J, Charles F, Chen M, Hatab SR. Antimicrobial Properties and Mechanism of Action of Some Plant Extracts Against Food Pathogens and Spoilage Microorganisms. Front Microbiol. 2018 Jul 24;9:1639. doi: 10.3389/fmicb.2018.01639.
- Shubha HS, Hiremath RS. Evaluation of antimicrobial activity of Rasaka Bhasma. Ayu. 2010 Apr;31(2):260-2. doi: 10.4103/0974-8520.72412.